Cohen ,M. M., Neri, G. &, Weksberg , R. (2002). Fragile X Syndrome. In M. M.Cohen, G. Neri &, R. Weksberg (Eds.), *Overgrowth Syndromes* (152-160). Oxford: Oxford University Press.

Fragile X Syndrome

More than 125 X-linked mental retardation (XLMR) syndromes have been identified, and of these, fragile X syndrome is the most common (43,59). The syndrome is the prototype of an ever growing list of disorders with dynamic mutations that result from instability of triplet repeats. The mutant gene in fragile X syndrome is FMRI (OMIM 309550)^a and the repeated triplet is CGG (68). Several excellent reviews are available, the one by Nussbaum and Ledbetter (44) being particularly exhaustive, and many international fragile X workshops have been held (21).

PREVALENCE

Prevalence estimates are summarized in Table 13–1. Fragile X syndrome is not as prevalent as initially estimated. The discrepancy can be explained by the use of a molecular diagnosis, which is more accurate than the cytogenetic test previously employed.

Prevalence estimates of healthy female carriers are also summarized in Table 13–1. The higher estimate in the study of Falik-Zaccai et al. (16) is partially attributable to the current lack of an accurate definition of a premutation (meiotically unstable allele). Further studies are needed to establish whether this unexpectedly high prevalence of premutation carriers is unique to the specific populations studied or whether it applies to other populations, which is probable (54). Evidence that expansion to full mutation on transmission from a premutated mother is more likely to occur in male fetuses than in female fetuses (34) and may explain a relative lack of premutated males in the general population (50). Large population studies on an unselected series of newborns would be useful in settling the true prevalence of affected (fully mutated) males, normal transmitting (premutated) males, and fully mutated and premutated female carriers. Caution must be exercised in planning such studies to avoid untoward effects on the screened subjects (42).

GENETICS

Many genetic and molecular papers have appeared (1,3,6,11,28,31,36,49,56,57,61,62). This section considers (a) gene structure and protein isoforms and (b) the origin and effects of full mutations.

Gene Structure and Protein Isoforms

The *FMR1* gene maps to Xq27.3 and has 17 exons spanning 38 kb of genomic DNA. The polymorphic CGG repeat is located in the 5' untranslated region of exon 1 and is included in all *FMR1* transcripts (3). *FMR1* was shown to be ubiquitously transcribed during murine and human embryogenesis with the highest levels of differentiated neurons in the hippocampus and basal ganglia (1); in adult mice, it is expressed only in neurons and spermatogonia. The 4.4 kb full-length mRNA encodes a protein with a max-

^aOn-Line Mendelian Inheritance in Man number.

<i>Table 13–1</i> Prevalence Estimates ^a of Fragile X	
Syndrome and Healthy Female Carriers	

Estimates	Reference
FRAGILE X SYNDROME	
67/100,000 25/100,000 ^b 17/100,000 ^b	Webb et al. (69) Morton et al. (40) de Vries et al. (14)
HEALTHY FEMALE CARRIERS	S
386/100,000 407/100,000° 1176/100,000°	Rousseau et al. (51) Ryynänen et al. (52) Falik-Zaccai et al. (16)

^aEstimates have been converted to standard epidemiologic form. With a standardized denominator, numerators can be compared more effectively.

^bMore realistic estimates by the use of molecular diagnosis, which is more accurate than the cytogenetic test previously employed.

 $^{\rm c}{\rm Difference}$ probably reflects current lack of an accurate definition of a premutation.

imum size of 632 amino acids and a molecular weight of 70–80 kDa (15). Although 20 different transcripts can be produced by alternative splicing (3), only 4 or 5 of them and their corresponding protein products are actually detected in various tissues. Isoform 7 (ISO7), which lacks only the 21 amino acids of exon 12, makes almost all the FMR1 protein and corresponds to the highest band on Western blotting with an approximate molecular weight of 80 kDa (57).

Two KH domains (KH1 and KH2) and one RGG box, common to several RNA-binding proteins, have been identified in exons 8, 10, and 15, respectively (56). It has been shown that FMR1 can bind synthetic RNAs in vitro, and the importance of KH domains was underscored by the description of a severely retarded fragile X patient with a point mutation change in a highly conserved KH2 isoleucine to asparagine (Ile304Asn) (11), which impaired the RNA binding activity of FMR1. It has also been shown that ISO7 is localized in the cytoplasm (15). In contrast, minor isoforms are lacking exon 14 and, with a different C-terminus (ISO6 or ISO12), are confined to the nucleus (57). Although no specific nuclear localization signal (NLS) is present in the first half of FMR1, studies with deletion constructs indicate that the N-terminus of the protein (exons 1-5) can direct the protein to the nucleus; sequences in exon 14 are essential for its subsequent export to the cytoplasm (57).

Origin and Effects of Full Mutations

In more than 95% of cases, fragile X syndrome is caused by a single type of mutation expansion and hypermethylation of a potentially unstable CGG trinucleotide repeat in the 5' UTR of the *FMR1* gene (full mutation). This causes transcriptional silencing of the gene and absence of the FMR1 protein. A full mutation appears to be generated by a lengthy multistep process requiring sequential action by a different mechanism (6). To date, no direct conversion of a wild-type to a fully mutated allele has been observed in fragile X families in which mothers of affected individuals were found to be carriers of an already expanded CGG triplet.

Different alleles at the CGG repeat are generally included in one of three classes depending on their total length: wild-type (6–50 repeats), premutation (50–200 repeats), and full mutation (200–1000 hypermethylated repeats). However, it has been shown that the boundaries between these classes are not absolute and that the initial instability depends not only on the total length but also on the repeat configuration. Detailed analysis of over 400 wild-type alleles has shown that the CGG repeat stretch is commonly interrupted by AGG triplets, mainly two, at intervals of 9–10 CGG units, which apparently have a stabilizing effect by preventing replication slippage.

The secondary structures formed by the full mutation and/or the increased number of CpG dinucleotide targets presumably favor the hypermethylation of the FMR1 gene harboring a full mutation (28).

The question of when and in which cells the pre-to-full expansion takes place is still debated. It was initially proposed that expansion to full mutation was postzygotic because sperm cells obtained from male patients were shown to harbor only premutations (49). However, recent evidence demonstrates the presence of full mutations in the oogonia and spermatogonia of female fetuses and male fetuses, respectively (36), supporting the hypothesis of a selection process that favors premutations in the male germline.

CLINICAL PHENOTYPE

General Features and Variability

The phenotype is variable. In newborns, the birthweight may be elevated, relative macrocephaly may be found, and the anterior fontanelle may be large. Typical adult males have tall stature, large testes, relative macrocephaly, prominent forehead, hypoteloric sunken eyes, long narrow face, midface hypoplasia, prominent mandible, and large ears (Figs. 13-1 to 13-3). The phenotype tends to change with age. In infants and adolescents, the phenotype is milder and, often, macrocephaly, increased height, and hypotonia are the only findings. Hypotonia is virtually a constant feature and is usually accompanied by joint laxity. Mitral valve prolapse is frequently observed in hemizygotes after 18 years of age. On occasion,

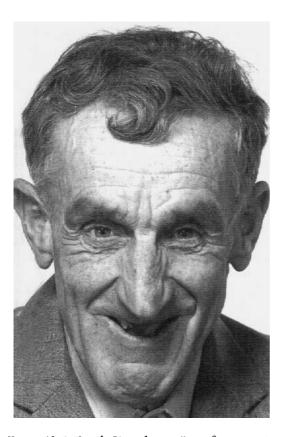


Figure 13–1 Fragile X syndrome. Long face, prominent forehead, sunken eyes, broad based nose, prominent chin, and prominent ears. From McDermott et al. (39).

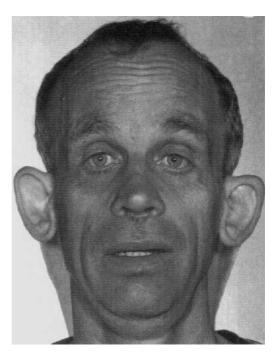


Figure 13–2 Fragile X syndrome. Narrow face and prominent ears. From McDermott et al. (39).

a small child may have the same phenotype as that found in typical adults.^b A subgroup of patients have been noted to have short stature and obesity. Infrequently, hemizygotes have been noted to appear entirely normal. About onethird of female carriers with the full mutation have a behavioral phenotype that includes mild mental retardation, shyness, poor eye contact, and learning disabilities. Some, particularly the more retarded ones, have the typical long face, mandibular prognathism, and large everted ears (10,19,20,22,30,33,39,58,59a).

Typical Craniofacial Appearance

In typical adult males, an increased head circumference and dolichocephaly are found together with a high quadrangular forehead, prominent supraorbital ridges, and a long narrow face. Puffiness under the eyes occurs in

^bParvari et al. (45) reported an 8.5 Mb deletion, including the *FMR1* gene, in a 4-year-old boy with a facial appearance consistent with fragile X syndrome and a height and weight above the 97th centile.



Figure 13–3 Fragile X syndrome. Macroorchidism. From McDermott et al. (39).

some cases. The palpebral fissure length is increased and the inter-inner canthal distance is decreased. Strabismus and refractive errors may be present. The nose often appears broadly based and midface hypoplasia is found (Figs. 13–1 and 13–2). The chin becomes long and prominent during adolescence. The ears are large, outstanding, and may be somewhat soft. The helices may be simple and the lobules may be absent. Otitis media is common and the palate is highly arched (20,39).

Central Nervous System and Performance

Central nervous system findings and performance have been discussed by many authors (9,10,19,20,22,30,33,41,46–48,53,59a,63,67,75). IQs in hemizygotes have been reported to range from 20–70. IQs in females with the full mutation range from 50–107. Delayed speech is very common. Retardation seems to increase with age. Those with higher IQs exhibit dysfluencies and stuttering and, frequently, a characteristic intonation. With lower IQs, less verbal ability is found together with repetitive phrases. Females who carry the full mutation may have learning disabilities. Even psychosis has been noted in some cases (9,19,20,22,53,63,75).

Neuropsychiatric evaluation shows delayed milestones, hypotonia, increased deep tendon reflexes, emotional instability, and automutilation, particularly handbiting. An adverse response to touch on the skin is found. Aggressive behavior occurs in about 50%. Seizures may be found during infancy; a characteristic EEG pattern has been reported as trains of medium-high voltage spikes discharging from the temporal region during sleep. A cognitive profile has been reported to consist of attention deficit hyperactivity disorder, oppositional defiant disorder, enuresis, and encopresis. Fragile X boys show greater variability in total sleep time than normal and difficulty in sleep maintenance compared with controls. In some cases, autism is found, characterized by avoidance of eye contact, hand flapping, or other stereotypic movements, perseverative speech, and echolalia (4, 19, 20, 22, 23, 30, 41, 67).

MRI of the brain shows volume conservation of tissue, diminished white-to-gray matter ratio, relatively enlarged caudate nucleus and hippocampus, and increased cerebrospinal fluid, particularly in the lateral ventricles. In males, a relative decrease in the size of the superior temporal gyrus and cerebellar vermis occurs with a relative increase in the size of the fourth ventricle. Likewise, in young females, a relative decrease in the size of the cerebellar vermis is found together with an increase in the size of the fourth ventricle. Neuropathologic studies have demonstrated more long dendritic spines and fewer shorter ones than normal with significant immature morphology in both temporal and visual cortical areas (29,46-48).

Connective Tissue Findings

Joint laxity involves particularly the thumbs but also the fingers, metacarpophalangeal joints, knees, and ankles. The skin feels velvety and soft over the dorsa of the hands. Single palmar creases are found in about one-fourth of the cases. Flat feet are common. Mitral valve prolapse, encountered in approximately 80% of hemizygotes after 18 years of age, is accompanied by aortic dilatation in about 15% of the cases (27).

Genitourinary System

Macro-orchidism, either unilateral or bilateral, is easier to detect in postpubertal males (Fig. 13–3). The testes tend to be softer than normal and the scrotum may be hyperpigmented. There is increased seminiferous tubule length and interstitial edema. The penis is often enlarged (9,14,20,32).

Early menopause and an increased rate of twinning, both reported in premutation carriers, are indications of ovarian failure, which has been found in about 20% of carriers. The data supporting increased twinning conflict and need to be resolved (19,33,55).

DIAGNOSIS

Molecular diagnosis of the CGG amplification is available since the cloning of the *FMR1* gene in 1991; it relies on Southern blotting and hybridization of specific probes, while PCR is employed to detect repeat lengths in the premutation range. The cytogenetic test in low-folate media is obsolete and has been abandoned. A rapid method based on antibody detection of the FMR1 protein in the cells of blood smears is useful for screening affected males (73,74) and has been adapted for uncultured amniotic cells (72) and hair roots (65,71,74).

Prenatal diagnosis depends on the availability of sufficient DNA to perform Southern blotting after double digestion including a methylationsensitive enzyme (usually EagI or BssHII), while the sex of the fetus can be determined by standard karyotype or Y-specific PCR analysis. False positives, due to suboptimal amplification, and false negatives, due to the possible presence of reverted alleles in the wild-type range, can occur when performing PCR alone on a sample from a male fetus. Furthermore, only direct DNA analysis after digestion with methylationsensitive enzymes can demonstrate the methylation status of the *FMR1* CpG island, especially in the presence of a full mutation (58). Given that in extra-embryonic tissues, such as chorionic villi, a full mutation may remain largely undermethylated until 10-11 weeks of gestation (5,35,60), chorionic villus sampling might not reveal the hypermethylation already present in the

embryonic tissues and may need confirmation by amniocentesis.

DIFFERENTIAL DIAGNOSIS

A subgroup of fragile X patients have short stature and obesity with a superficial resemblance to Prader-Willi syndrome (12). Several patients initially thought to have Sotos syndrome were diagnosed as having fragile X syndrome. Pitfalls in clinical diagnosis justify the view that every mentally retarded patient should be tested for fragile X syndrome in the absence of another reasonable diagnosis. Difficulties in the diagnosis of fragile X syndrome in young children have been discussed by Stoll (59a).

GUIDELINES FOR HEALTH SUPERVISION

Useful guidelines for health supervision of fragile X children have been published by the American Academy of Pediatrics (2) and include advice for both physical and behavioral aspects of the syndrome. After confirming the diagnosis with molecular testing and appropriate parental counseling for recurrence risk for subsequent pregnancies, a series of medical examinations can be envisioned depending on the age of the child. Development during the first year of life can be normal, although hypotonia and irritability may be apparent. In early childhood, the following examinations are important: ophthalmologic examination (strabismus, myopia), echocardiogram if a murmur or click is present (mitral valve prolapse), and orthopedic examination (flat feet, scoliosis, and loose joints). Inguinal hernias should also be excluded. A history of seizures or startle episodes should be reviewed; performing an EEG may be appropriate, although antiepileptic medication after a single seizure is not advisable, given the self-limited course of epileptic manifestations in adolescence (41). Hyperactive behavior and severe attention deficit, which are a major concern during school age, can be treated pharmacologically (24). Torrioli et al. (64) reported the results of a preliminary clinical trial, suggesting that L-acetylcarnitine might be beneficial in ameliorating the hyperactive behavior of affected children. However, socialization and school integration, possibly within a mainstream program with individual support, are important in helping to overcome these problems. Sports and regular physical activity are important to counteract hypotonic posture and to improve motor coordination. Speech, language, and occupational therapy should be goal oriented to help adolescents and young adults attain as much autonomy as possible. Support from family organizations is important, especially for parents and sibs.

Treatments aimed at recovering the function of the FMR1 gene have been attempted with folic acid because of its action on cytogenetic expression of the fragile site. Although some reports indicated some beneficial effects on behavior (15), others have not confirmed these observations (18,70). Folate supplementation has no efficacy for the treatment of fragile X syndrome patients. Observations on intellectually normal (58) or minimally affected (25,37) males with an unmethylated full mutation confirm that the abnormally amplified CGG tract per se can still be transcribed and translated. Even if translation may not be completely efficient (17), lymphoblastoid cell lines containing only unmethylated full mutations of two such males have shown the presence of FMR1 protein in every cell, but at reduced levels (58). Given the observations of these exceptional individuals and knowing that the coding sequence of the mutated FMR1 gene is intact, Chiurazzi et al. (7) tested the possibility of restoring its activity in vitro, employing a DNA demethylation protocol. These authors obtained in vitro reactivation of FMR1 expression after inducing DNA demethylation with 5-azadeoxycytidine in lymphoblastoid cell lines from patients.

Given that the effect of DNA hypermethylation in silencing the *FMR1* gene seems to be potentiated by deacetylation of histones, Chiurazzi et al. (8) performed another set of experiments with phenylbutyrate, an inhibitor of histone deacetylase, and showed that treatment of fragile X cell lines restores FMR1 transcription. Although the effect was weak, it appeared to be synergistic with that of 5-azadeoxycytidine. Phenylbutyrate is structurally similar to L-acetylcarnitine, used in vivo by Torrioli et al. (64) in the clinical trial discussed above. McConkie-Rosell et al. (38) carried out a longitudinal study of women-at-risk for inheriting the fragile X mutation. Problems addressed were (a) how upsetting the women perceived their information to be, (b) how serious a problem they perceived fragile X syndrome to be, and (c) their feelings about the carrier testing process. Such information is useful for counseling purposes.

REFERENCES

- Abitbol M, Menini C, Delezoide AL, Rhyner T, Vekemans M, Mallet J: Nucleus basalis magnocellularis and hippocampus are the major sites of FMR1 expression in the human fetal brain. Nat Genet 4:147–153, 1993.
- 2. American Academy of Pediatrics, Committee on Genetics: Health supervision for children with fragile X syndrome. Pediatrics 98:297–300, 1996.
- 3. Ashley CT, Sutcliffe JS, Kunst CB, Leiner HA, Eichler EE, Nelson DL, Warren ST: Human and murine *FMR1*: alternative splicing and translational initiation downstream of the CGG repeat. Nat Genet 4:244–251, 1993.
- Backes M, Genç B, Schreck J, Doerfler W, Lehmkuhl C, von Gontard A: Cognitive and behavioral profile of fragile X boys: Correlations to molecular data. Am J Med Genet 95:150–156, 2000.
- Castellvi-Bel S, Mila M, Soler A, Carrio A, Sanchez A, Villa M, Jimenez MD, Estivill X: Prenatal diagnosis of fragile X syndrome: (CGG)_n expansion and methylation of chorionic villus samples. Prenatal Diagn 15:801–807, 1995.
- Chiurazzi P, Macpherson J, Sherman S, Neri G: Editorial Comment: Significance of linkage disequilibrium between the fragile X locus and its flanking markers. Am J Med Genet 64:203–208, 1996.
- Chiurazzi P, Pomponi MG, Willemsen R, Oostra BA, Neri G: In vitro reactivation of the *FMR1* gene involved in fragile X syndrome. Hum Molec Genet 7:109–113, 1998.
- 8. Chiurazzi P, Pomponi MG, Pietrobono R, Bakker CE, Neri G, Oostra B: Synergistic effect of histone hyperacetylation and DNA demethylation in the reactivation of the *FMR1* gene. Hum Molec Genet 8:2317–2323, 1999.
- 9. Chudley AE, Hagerman RJ: Fragile X syndrome. J Pediatr 110:821-831, 1987.
- Cianchetti C, Sannio-Fancello G, Fratta A-L, Manconi F, Orano A, Pischedda M-P, Pruna D, Spinicci G, Archidiacono N, Filippi G: Neuropsychological, psychiatric, and physical manifestations in 149 members from 18 fragile X families. Am J Med Genet 40:234–243, 1991.

- 11. De Boulle K, Verkerk AJMH, Reyniers E, Vits L, Hendrickx J, Van Ro B, van den Bos F, de Graaff E, Oostra BA, Willems PJ: A point mutation in the *FMR1* gene associated with fragile X mental retardation. Nat Genet 3:31–35, 1993.
- de Vries BB, Fryns JP, Butler MG, Canziani F, Wesby-van-Swaay E, van-Hemel JO, Oostra BA, Halley DJ, Niermeijer MF: Clinical and molecular studies in fragile X patients with a Prader-Willi-like phenotype. J Med Genet 30:761–766, 1993.
- de Vries BBA, Robinson H, Stolte-Dijkstra F, Tjon Pian Gi CV, Dijkstra PF, van Doorn J, Halley DJ, Oostra BA, Turner G, Niermeijer MF: General overgrowth in the fragile X syndrome: Variability in the phenotype expression of the *FMR1* gene mutation. J Med Genet 32:764–769, 1995.
- 14. de Vries BBA, van den Ouweland AMW, Mohkamsing S, Duivenvoorden HJ, Moi E, Gelsema K, van Rijn M, Halley DJ, Sandkuijl LA, Oostra BA, Tibben A, Niermeijer MF: Screening and diagnosis of the fragile X syndrome among the mentally retarded: An epidemiological and psychological survey. Am J Hum Genet 61:660–667, 1997.
- Devys D, Lutz Y, Rouyer N, Bellocq JP, Mandel JL: The FMR1 protein is cytoplasmic, most abundant in neurons and appears normal in carriers of a fragile X premutation. Nat Genet 4:335–340, 1993.
- Falik-Zaccai TC, Shachak E, Borochowits Z, Magal N, Zatz S, Schochat M, Ziu H, Navon R, Legum C, Shomrat R: Fragile X syndrome: Population carrier screening and implication for prenatal diagnosis. Am J Hum Genet 65:A214, 1999.
- Feng Y, Zhang F, Lokey LR, Chastain JL, Lakkis L, Eberhart D, Warren ST: Translational suppression by trinucleotide repeat expansion at FMR1. Science 268:731–734, 1995.
- Froster-Iskenius U, Bodeker R, Oepen T, Matthes R, Piper U, Schwinger E: Folic acid treatment in males and females with fragile-(X)syndrome. Am J Med Genet 23:273–289, 1986.
- 19. Fryns J-P: The female and the fragile X: A study of 144 obligate female carriers. Am J Med Genet 23:157–169, 1986.
- Fryns J-P: The fragile X syndrome: A study of 83 families. Clin Genet 26:497–528, 1984.
- Fryns J-P, Borghgraef M, Brown TW, Chelly J, Fisch GS, Hamel B, Hanauer A, Lacombe D, Luo L, MacPherson JN, Mandel J-L, Moraine C, Mulley J, Nelson D, Oostra B, Partington M, Ramakers GJA, Ropers H-H, Rousseau F, Schwartz C, Steinbach P, Stoll C, Tranebjaerg L, Turner G, Van Bokhoven H, Vianna-Morgante A, Villard L, Warren ST: 9th international workshop on fragile X syndrome and X-linked mental retardation. Am J Med Genet 94:345– 360, 2000.

- Fryns J-P, Jacobs J, Kleczkowska A, van den Berghe H: The psychological profile of the fragile X syndrome. Clin Genet 25:131–134, 1984.
- Gould EL, Loesch DZ, Martin MJ, Hagerman RJ, Armstrong SM, Huggins RM: Melatonin profiles and sleep characteristics in boys with fragile X syndrome: A preliminary study. Am J Med Genet 95:307–315, 2000.
- Hagerman R: Fragile X: treatment of hyperactivity. Pediatr 99:753, 1997.
- 25. Hagerman RJ, Hull CE, Safanda JF, Carpenter L, Staley LW, O'Connor RA, Seydel C, Mazzocco M, Snow K, Thibodeau SN, Kuhl D, Nelson DL, Caskey CT, Taylor AK: High functioning fragile X males: Demonstration of an unmethylated fully expanded *FMR1* mutation associated with protein expression. Am J Med Genet 51:298–308, 1994.
- Hagerman RJ, Jackson AW, Levitas A, Braden M, McBogg P, Kemper M, McGavran L, Berry R, Matus I, Hagerman PJ: Oral folic acid versus placebo in the treatment of males with the fragile X syndrome. Am J Med Genet 23:241–262, 1986.
- Hagerman RJ, Van Housen K, Smith ACM, Mc-Gavran L: Consideration of connective tissue dysfunction in the fragile X syndrome. Am J Med Genet 17:111–121, 1984.
- Hansen RS, Canfield TK, Lamb MM, Gartler SM, Laird CD: Association of fragile X syndrome with delayed replication of the *FMR1* gene. Cell 73:1403–1409, 1993.
- 29. Irwin SA, Patel B, Idupulapati M, Harris JB, Crisostomo RA, Larsen BP, Kooy F, Willems PJ, Cras P, Kozlowski PB, Swain RA, Weiler IJ, Greenough WT: Abnormal dendritic spine characteristics in the temporal and visual cortices of patients with fragile-X syndrome: A quantitative examination. Am J Med Genet 98:161–167, 2001.
- Lachiewicz AM, Dawson DV, Spiridigliozzi GA: Physical characteristics of young boys with fragile X syndrome: Reasons for difficulties in making a diagnosis in young males. Am J Med Genet 92:229–236, 2000.
- Laggerbauer B, Ostareck D, Keidel E-M, Ostareck-Lederer A, Fischer U: Evidence that fragile X mental retardation protein is a negative regulator of translation. Hum Mol Genet 10:329–338, 2001.
- Limprasert P, Jarvratanasirikul S, Vasiknanonte P: Letter to the Editor: Unilateral macroorchidism in fragile X syndrome. Am J Med Genet 95:516–517, 2000.
- Loesch DZ, Hay DA: Clinical features and reproductive patterns in fragile X female heterozygotes. J Med Genet 25:407–414, 1988.
- 34. Loesch DZ, Huggins R, Petrovic L, Slater H: Expansion of the CGG repeat in fragile X in the FMR1 gene depends on the sex of the offspring. Am J Hum Genet 57:1408–1413, 1995.

- 35. Luo S, Courtland Robinson J, Reiss AL, Migeon BR: DNA methylation of the fragile X locus in somatic and germ cells during fetal development: Relevance to the fragile X syndrome and X inactivation. Somat Cell Molec Genet 19:393– 404, 1993.
- Malter HE, Iberr C, Willemsen R, de Graaff E, Tarleton JC, Leisti J, Warren ST, Oostra BA: Characterization of the full fragile X syndrome mutation in fetal gametes. Nat Genet 15:165– 169, 1997.
- 37. McConkie-Rosell A, Lachiewicz AM, Spiridigliozzi GA, Tarleton J, Schoenwald S, Phelan MC, Goonewardena P, Ding X, Brown WT: Evidence that methylation of the FMR-1 locus is responsible for variable phenotypic expression of the fragile X syndrome. Am J Hum Genet 53:800– 809, 1993.
- McConkie-Rosell A, Spiridigliozzi GA, Sullivan JA, Dawson DV, Lachiewicz AM: Longitudinal study of the carrier testing process for fragile X syndrome: Perceptions and coping. Am J Med Genet 98:37–45, 2001.
- McDermott A: Fragile X chromosome: Clinical and cytogenetic studies from seven families. J Med Genet 20:169–171, 1983.
- Morton JE, Bundey S, Webb TP, MacDonald F, Rinde PM, Bullock S: Fragile X syndrome is less common than previously estimated. J Med Genet 34:1–5, 1997.
- Musumeci SA, Ferri R, Elia M, Colognola RM, Bergonzi P, Tassinari CA: Epilepsy and fragile X syndrome: a follow-up study. Am J Med Genet 38:511–513, 1991.
- Neri G, Chiurazzi P: Fragile X syndrome screening: A current opinion. Commun Genet 3:38–40, 2000.
- 43. Neri G, Opitz JM: Sixty years of X-linked mental retardation: A historical footnote. Am J Med Genet 97:228–233, 2000.
- Nussbaum RL, Ledbetter DH: The fragile X syndrome. In *The Metabolic and Molecular Bases of Inherited Disease*. CR Scriver, AL Beaudet, WS Sly, D. Valle D, eds., Chapter 19, pp. 795–810, 1995.
- 45. Parvari R, Mumm S, Galil A, Manor E, Bar-David Y, Carmi R: Deletion of 8.5 Mb, including the *FMR1* gene, in a male with the fragile X syndrome phenotype and overgrowth. Am J Med Genet 83:302–307, 1999.
- Reiss AL, Abrams MT, Greenlaw R, Freund L, Denckla MB: Neurodevelopmental effects of the *FMR1* full mutation in humans. Nat Med 1:159–167, 1995.
- Reiss AL, Aylward E, Freund LS, Joshi PK, Bryan RN: Neuroanatomy of fragile X syndrome: The posterior fossa. Ann Neurol 29:26– 32, 1991.
- Reiss AL, Lee J, Freund L: Neuroanatomy of fragile X syndrome: The temporal lobe. Neurology 44:1317–1324, 1994.

- 49. Reyniers E, Vits L, De Boulle K, van Velzen D, de Graaff E, Verkerk AJMH, Jorens HZ, Darby JK, Oostra BA, Willems PJ: The full mutation in the *FMR1* gene of male fragile X patients is absent in their sperm. Nat Genet 4:143–146, 1993.
- 50. Rousseau F, Morel ML, Rouillard P, Khandjian EW, Morgan K: Surprisingly low prevalence of the FMR1 premutations among males from the general population. Am J Hum Genet 59 (Suppl): A188, 1996.
- Rousseau F, Rouillard P, Morel ML, Khandjian EW, Morgan K: Prevalence of carriers of premutation-size alleles of the *FMR1* gene and implications for the population genetics of the fragile X syndrome. Am J Hum Genet 57:1006– 1018, 1995.
- Ryynänen M, Heinonen S, Makkonen M, Kajanoja E, Mannermaa A, Pertti K: Feasibility and acceptance of screening for fragile X mutations in low-risk pregnancies. Eur J Hum Genet 7:212–216, 1999.
- 53. Schapiro MB, Murphy DGM, Hagerman RJ, Azari NP, Alexander GE, Miezejeski CM, Hinton VJ, Horwitz B, Haxby JV, Kumar A, White B, Grady CL: Adult fragile X syndrome: Neuropsychology, brain anatomy, and metabolism. Am J Med Genet 60:480–493, 1995.
- 54. Sherman SL: The high prevalence of fragile X premutation carrier females: is this frequency unique to the French Canadian population? Am J Hum Genet 57:991–993, 1995.
- Sherman SL: Premature ovarian failure in the fragile X syndrome. Am J Med Genet 97:189– 194, 2000.
- Siomi H, Siomi MC, Nussbaum RL, Dreyfuss G: The protein product of the fragile X gene, *FMR1*, has characteristics of an RNA-binding protein. Cell 74:291–298, 1993.
- Sittler A, Devys D, Weber C, Mandel JL: Alternative splicing of exon 14 determines nuclear or cytoplasmic localisation of FMR1 protein isoforms. Hum Molec Genet 5:95–102, 1996.
- Smeets HJM, Smits APT, Verheij C, Theelen JPG, van de Burgt I, Hoogeven AT, Oosterwijk JC, Oostra BA: Normal phenotype in two brothers with a full FMR1 mutation. Hum Mol Genet 4:2103–2108, 1995.
- Stevenson RE: Splitting and lumping in the nosology of XLMR. Am J Med Genet 97:174– 182, 2000.
- 59a. Stoll C: Problems in the diagnosis of fragile X syndrome in young children are still present. Am J Med Genet 100:110–115, 2001.
- 60. Sutcliffe JS, Nelson DL, Zhang F, Pieretti M, Caskey CT, Saxe D, Warren ST: DNA methylation represses FMR-1 transcription in fragile X syndrome. Hum Mol Genet 1:397–400, 1992.
- Tassone F, Hagerman RJ, Chamberlain WD, Hagerman PJ: Transcription of the *FMR1* gene in individuals with fragile X syndrome. Am J Med Genet 97:195–203, 2000.

- 62. Tassone F, Hagerman RJ, Loesch DZ, Lachiewicz A, Taylor AK, Hagerman PJ: Fragile X males with unmethylated, full mutation trinucleotide repeat expansions have elevated levels of FMR1 messenger RNA. Am J Med Genet 94:232–236, 2000.
- Theobald TM, Hay DA, Judge C: Individual variation and specific cognitive deficits in the fra(X) syndrome. Am J Med Cenet 28:1–11, 1987.
- 64. Torrioli MG, Vernacotola S, Mariotti P, Bianchi E, De Gaetano A, Calvani M, Chiurazzi P, Neri G: Double-blind, placebo-controlled study of L-acetylcarnitine for the treatment of hyperactive behavior in fragile X syndrome. Am J Med Genet 87:366–368, 1999.
- 65. Tunçbilek E, Alikasifoglu M, Aktas D, Duman F, Yanik H, Anar B, Oostra B, Willemsen R: Screening for the fragile X syndrome among mentally retarded males by hair root analysis. Am J Med Genet 95:105–107, 2000.
- 66. Van Roy BC, De Smedt MC, Raes RA, Dumon JE, Leroy JG: Fragile X trait in a large kindred: Transmission also through normal males. J Med Genet 20:286–289, 1983.
- 67. Veenema H, Veenema T, Geraedts J: The fragile X syndrome in a large family. II. Psy-chological investigation. J Med Genet 24:32–38, 1987.
- 68. Verkerk AJMH, Pieretti M, Sutcliffe JS, Su YH, Kuhl DPA, Pizzuti A, Reiner O, Richards S, Victoria MF, Zhang F, Eussen BE, van Ommen GJB, Blonden LAJ, Riggins GJ, Chastain JL, Kunst CB, Galjaard H, Caskey CT, Nelson DL,

Oostra BA, Warren ST: Identification of a gene (FMR-1) containing a CGG repeat coincident with a breakpoint cluster region exhibiting length variation in fragile X syndrome. Cell 65:905–914, 1991.

- Webb TP, Bundey SE, Thake AI, Todd J: Population incidence and segregation ratios in the Martin-Bell syndrome. Am J Med Genet 23: 573–580, 1986.
- Webb T, Crawley P, Bundey S: Folate treatment of a boy with fragile-X syndrome. J Ment Defic Res 34:67-73, 1990.
- Willemsen R, Anar B, de Diego Otero Y, de Vries BB, Hilhorst-Hofstee Y, Smits A, van Looveren E, Willems PJ, Galjaardt H, Oostra BA: Non invasive test for fragile X syndrome, using hair root analysis. Am J Hum Genet 65:98–103, 1999.
- 72. Willemsen R, Los F, Mohkamssing S, Van den Ouweland A, Deelen W, Galjaard H, Oostra BA: Rapid antibody test for prenatal diagnosis of fragile X syndrome on amniotic fluid cells: A new appraisal. J Med Genet 34:250–251, 1997.
- Willemsen R, Mohkamsing S, De Vries B, Devys D, van den Ouweland A, Mandel JL, Galjaard H, Oostra BA: Rapid antibody test for fragile X syndrome. Lancet 345:1147–1148, 1995.
- Willemsen R, Oostra BA: FMRP detection assay for the diagnosis of the fragile X syndrome. Am J Med Genet 97:183–188, 2000.
- Wisniewski KE, Segan SM, Miezejeski CM, Sersen EA, Rudelli RD: The fra(X)syndrome: Neurological, electrophysiological, and neuropathological abnormalities. Am J Med Genet 38:476–480, 1991.